Acute Infectious Diarrhea

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An otherwise healthy 23-year-old man presents after the acute onset of watery diarrhea that has persisted for two days. He reports associated nausea and cramping but no emesis and is febrile, with a temperature of 38.7°C (101.7°F). How should he be evaluated and treated?

Despite reductions in mortality worldwide, diarrhea still accounts for more than 2 million deaths annually and is associated with impaired physical and cognitive development in resource-limited countries. In the United States, an estimated 211 million to 375 million episodes of acute diarrhea occur each year (1.4 episodes per person per year); such episodes are responsible for more than 900,000 hospitalizations and 6000 deaths annually.

Acute diarrhea, defined as an increased frequency of defecation (three or more times per day or at least 200 g of stool per day) lasting less than 14 days, may be accompanied by nausea, vomiting, abdominal cramping, clinically significant systemic symptoms, or malnutrition. We focus here on acute infectious diarrhea in immunocompetent adults in industrialized countries.

Strategies and Evidence

Microbiology
The Foodborne Diseases Active Surveillance Network (FoodNet) of the Centers for Disease Control and Prevention (CDC) collects data on the incidence of diarrhea attributable to nine enteropathogens in 13 percent of the U.S. population (37.4 million people) living in nine states. Of these, the pathogens responsible for the most cases of diarrhea in 2002 were salmonella (16.1 cases per 100,000 population), campylobacter (13.4 cases per 100,000 population), shigella (10.3 cases per 100,000 population), Escherichia coli O157:H7 (1.7 cases per 100,000 population), and cryptosporidium (1.4 cases per 100,000 population); vibrio, yersinia, listeria, and cyclospora were reported in fewer than 1 person per 100,000. Other enteropathogens for which diagnostic testing is readily available include Clostridium difficile, giardia, rotavirus, and Entamoeba histolytica. Additional agents of infectious diarrhea for which clinical diagnostic testing is not routinely available include enterotoxigenic, enteropathogenic, enteroaggregative, and enteroinvasive strains of E. coli, toxin-producing Clostridium perfringens, Staphylococcus aureus, Bacillus cereus, and noroviruses.

Evaluation
Thorough clinical evaluation of a patient who presents with acute diarrhea is essential in order to guide a cost-effective, evidence-based approach to initial diagnostic test-
ing and therapy. In six studies conducted between 1980 and 1997, the diagnostic yield of stool cultures ranged from 1.5 to 5.6 percent.6 The estimated cost of $952 to $1,200 per positive culture can be reduced through improved selection and testing of the specimens submitted. As is indicated in Figure 1, the initial clinical evaluation of the patient with acute diarrhea should focus on the assessment of the severity of the illness, the need for rehydration, and the identification of likely causes on the basis of the history and clinical findings.

Because most diarrheal illnesses are self-limited or viral, and nearly half last less than one day,3 microbiologic investigation is usually unnecessary for patients who present within 24 hours after the onset of diarrhea, unless such patients are dehydrated or febrile or have blood or pus in their stool. The characteristics of the illness, the epidemiologic setting, and the public health implications help to determine whether and what types of fecal testing are appropriate. The major epidemiologic and clinical features associated with common enteropathogens are listed in Table 1.

In the United States, most diarrheal illnesses occur during the winter months; these illnesses are commonly associated with noroviruses (in families and in outbreaks, including those in nursing homes)7 or rotaviruses (in young children). Since these illnesses are typically short-lived (lasting one to three days) and self-limited, supportive fluid therapy is usually sufficient. Systemic illness, fever, or bloody stools should prompt routine stool testing for salmonella, shigella, campylobacter, and (especially if there are bloody stools) Shiga toxin–producing E. coli. When diarrhea is persistent or recurring, or if the history (for instance, of fever or tenesmus) is equivocal, fecal testing by means of microscopy for polymorphonuclear leukocytes or by means of immunoassay for the neutrophil marker lactoferrin can quickly provide additional laboratory evidence to support a presumptive diagnosis of inflammatory diarrhea.

In equivocal cases, the negative predictive value of fecal lactoferrin testing may help to determine the need for routine bacteriologic culture for campylobacter, salmonella, and shigella.8 In developed countries, the sensitivity and specificity of fecal leukocytes for inflammatory diarrhea are 0.73 and 0.84, respectively, and the sensitivity and specificity of lactoferrin for inflammatory diarrhea are 0.92 and 0.79, respectively; in resource-poor countries, the ranges are broader.9

Other testing is necessary in special circumstanc-
APPROACHES TO THERAPY

Rehydration and Nutrition

Regardless of the causative agent, initial therapy should include rehydration. Unless the patient is comatose or severely dehydrated, oral rehydration with a glucose-based electrolyte solution is preferred. The standard formulation recommended by the World Health Organization (WHO) or a newer reduced-osmolarity formula for children can be lifesaving in resource-poor regions and is valuable in the industrialized world for infants, the elderly, immunocompromised patients, and anyone with profuse watery diarrhea (information is available on the Web at http://www.who.int/child-adolescent-health/New_Publications/CHILD_HEALTH/WHO_FCH_CAH_01.22.htm). Most adults in developed countries who have acute diarrhea, including travelers, should be encouraged to drink fluids and take in salt in soups and salted crackers. Nutritional support with continued feeding improves outcomes in children. The use of a “BRAT” diet (bananas, rice, applesauce, and toast) with avoidance of milk products (since a transient lactase deficiency may occur) is commonly recommended, although supporting data are limited.15

Nonspecific Symptomatic Therapy

Nearly 400 over-the-counter products are promoted in the United States for their antidiarrheal properties, but few have been demonstrated to be effective in randomized, controlled trials. Only in the cases of loperamide, bismuth subsalicylate, and kaolin has sufficient evidence of efficacy and safety been collected to permit labeling as an antidiarrheal agent.16

Loperamide, the antimitotic agent of choice for adults, inhibits intestinal peristalsis and has anti-secretory properties, but unlike other opiates (such as codeine, diphenoxylate, and paregoric), it does not penetrate the nervous system and has no substantial potential for addiction.17 When it is used with antibiotics for traveler’s diarrhea18 or bacillary dysentery,19 it may reduce the duration of diarrhea by as much as one day. Because antimotility agents have been implicated in prolonged fever in human volunteers with shigellosis,20 toxic megacolon in patients with C. difficile infection, and the hemolytic–uremic syndrome in children infected with Shiga toxin–producing E. coli,21 these agents should be avoided in patients with bloody diarrhea or suspected inflammatory diarrhea.

Although it has not been as effective as loperamide in direct comparisons,22 bismuth subsalicylate can alleviate stool output in children23 or the symptoms of diarrhea, nausea, and abdominal pain in patients with traveler’s diarrhea.24 Bismuth subsalicylate significantly decreased the median duration of experimental norovirus illness from 27 hours to 20 hours.25 There are insufficient data from clinical trials to support the use of adsorbents such as kaolin–pectin, activated charcoal, and attapulgite. Given the limited scientific evidence, limited clinical benefit, and concern about the toxicity of antidiarrheal agents, pediatric guidelines discourage their use in children.26,27

Antimicrobial Therapy

Whereas appropriate antibiotics are effective in the treatment of shigellosis, traveler’s diarrhea, C. difficile–associated diarrhea, and, if given early, campylobacteriosis, antibiotics may prolong the duration of shedding of salmonella28 or C. difficile29 and may increase the risk of life-threatening complications of Shiga toxin–producing E. coli infection.30 The clinical benefit should be weighed against the cost, the risk of adverse reactions, and the risk of harmful eradication of normal flora or the induction of Shiga-toxin production.31 Antibiotic therapies for specific pathogens are listed in Table 2.
Perform initial assessment
Dehydration
Duration (>1 day)
Inflammation (indicated by fever, presence of blood in stool, tenesmus)

Provide symptomatic treatment
Rehydration
Treatment of symptoms (if necessary, consider bismuth subsalicylate or loperamide if diarrhea is not inflammatory or bloody)

Stratify subsequent management according to clinical and epidemiologic features
Epidemiologic clues:
Food, antibiotics, sexual activity, travel, day-care attendance, other illnesses, outbreaks, season
Clinical clues:
Bloody diarrhea, abdominal pain, dysentery, wasting, fecal inflammation

Obtain fecal specimen for analysis if severe, bloody, inflammatory, or persistent diarrhea or if outbreak suspected:

<table>
<thead>
<tr>
<th>Community-acquired or traveler’s diarrhea</th>
<th>Nosocomial diarrhea (onset &gt;3 days after hospitalization)</th>
<th>Persistent diarrhea (&gt;7 days)</th>
<th>If patient is immunocompromised (especially if HIV+) add</th>
</tr>
</thead>
<tbody>
<tr>
<td>Culture or test for Salmonella, shigella, campylobacter + (if history of bloody diarrhea or hemolytic–uremic syndrome) E. coli O157:H7 + shiga-like toxin + (if recent antibiotics, chemotherapy, or hospitalization) C. difficile toxins A and B</td>
<td>Test for C. difficile toxins A and B + (if outbreak or if patient is &gt;65 yr of age with coexisting conditions, immunocompromised, or neutropenic or if systemic enteric infection is suspected) salmonella, shigella, campylobacter, + (if bloody diarrhea) shiga toxin–producing E. coli</td>
<td>Consider protozoa: Giardia, cryptosporidium, cyclospora, Isospora belli + screening for inflammation</td>
<td>Test for Microsporidia, Mycobacterium avium complex, cytomegalovirus</td>
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</tbody>
</table>

Consider antimicrobial therapy for specific pathogens

Report specific diarrheal diseases promptly to public health authorities
In suspected outbreaks, save culture plates and isolates and freeze fecal and food or water specimens at −70°C. Diarrheal diseases designated as notifiable at the national level in the United States include cholera, cryptosporidiosis, giardiasis, salmonellosis, shigellosis, and infection with shiga toxin–producing E. coli.
timicrobial therapy shortens the average duration of diarrhea by 2.4 days or more, decreases the duration of fever and tenesmus, and reduces the excretion of infectious organisms. Antimicrobial therapy is of greatest benefit when it is started early, and it probably reduces person-to-person spread. Increased resistance now limits the effectiveness of trimethoprim–sulfamethoxazole, ampicillin, and nalidixic acid, which were once considered optimal therapies for shigellosis. Clinical trials have established the efficacy of newer fluoroquinolones for adults with shigellosis, and accumulating data suggest that ciprofloxacin is also safe and effective in children with shigellosis.

In immunocompetent patients with campylobacteriosis, erythromycin reduces carriage of the organism but reduces diarrhea only when it is given within four days after the onset of symptoms. Immunosuppressed patients (particularly those who have the acquired immunodeficiency syndrome [AIDS] and those who are pregnant, who have an increased risk of complications of campylobacter

<table>
<thead>
<tr>
<th>Pathogen</th>
<th>Common Epidemiologic Settings or Modes of Transmission</th>
<th>Clinical Features</th>
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<tbody>
<tr>
<td>Salmonella</td>
<td>Outbreaks due to foodborne transmission, community-acquired</td>
<td>Fever: Common; Abdominal Pain: Common; Bloody Stool: Occurs; Vomiting, Nausea, or Both: Occurs; Fecal Evidence of Inflammation: Common; Hemepositive Stool: Variable</td>
</tr>
<tr>
<td>Campylobacter</td>
<td>Community-acquired, consumption of undercooked poultry</td>
<td>Fever: Common; Abdominal Pain: Common; Bloody Stool: Occurs; Vomiting, Nausea, or Both: Occurs; Fecal Evidence of Inflammation: Common; Hemepositive Stool: Variable</td>
</tr>
<tr>
<td>Shigella</td>
<td>Community-acquired, person-to-person</td>
<td>Fever: Common; Abdominal Pain: Common; Bloody Stool: Occurs; Vomiting, Nausea, or Both: Occurs; Fecal Evidence of Inflammation: Common; Hemepositive Stool: Variable</td>
</tr>
<tr>
<td>Shiga toxin–producing E. coli (including O157:H7)†</td>
<td>Outbreaks due to foodborne transmission, especially through ingestion of undercooked hamburger or raw seed sprouts</td>
<td>Fever: Atypical; Abdominal Pain: Common; Bloody Stool: Occurs; Vomiting, Nausea, or Both: Occurs; Fecal Evidence of Inflammation: Common; Hemepositive Stool: Variable</td>
</tr>
<tr>
<td>C. difficile‡</td>
<td>Nosocomial spread, antibiotic use</td>
<td>Fever: Occurs; Abdominal Pain: Occurs; Bloody Stool: NC; Vomiting, Nausea, or Both: NC; Fecal Evidence of Inflammation: Common; Hemepositive Stool: Occurs</td>
</tr>
<tr>
<td>Vibrio</td>
<td>Ingestion of seafood</td>
<td>Fever: Variable; Abdominal Pain: Variable; Bloody Stool: Variable; Vomiting, Nausea, or Both: Variable; Fecal Evidence of Inflammation: Variable; Hemepositive Stool: Variable</td>
</tr>
<tr>
<td>Yersinia</td>
<td>Community-acquired, foodborne transmission</td>
<td>Fever: Common; Abdominal Pain: Occurs; Bloody Stool: Occurs; Vomiting, Nausea, or Both: Occurs; Fecal Evidence of Inflammation: Occurs; Hemepositive Stool: Occurs</td>
</tr>
<tr>
<td>E. histolytica§</td>
<td>Travel to tropical regions, recent emigration from such regions</td>
<td>Fever: Occurs; Abdominal Pain: Variable; Bloody Stool: Variable; Vomiting, Nausea, or Both: Variable; Fecal Evidence of Inflammation: Variable; Hemepositive Stool: Common</td>
</tr>
<tr>
<td>Cryptosporidium¶</td>
<td>Outbreaks due to waterborne transmission, travel, immunocompromised hosts</td>
<td>Fever: Variable; Abdominal Pain: Variable; Bloody Stool: NC; Vomiting, Nausea, or Both: NC; Fecal Evidence of Inflammation: Occurs; Hemepositive Stool: None to mild</td>
</tr>
<tr>
<td>Cyclospora¿</td>
<td>Outbreaks due to foodborne transmission, travel</td>
<td>Fever: Variable; Abdominal Pain: Variable; Bloody Stool: NC; Vomiting, Nausea, or Both: NC; Fecal Evidence of Inflammation: NC; Hemepositive Stool: NC</td>
</tr>
<tr>
<td>Giardia</td>
<td>Day care, outbreaks due to waterborne transmission, IgA deficiency</td>
<td>Fever: NC; Abdominal Pain: Common; Bloody Stool: NC; Vomiting, Nausea, or Both: NC; Fecal Evidence of Inflammation: NC; Hemepositive Stool: NC</td>
</tr>
<tr>
<td>Norovirus**</td>
<td>Winter outbreaks of vomiting or diarrhea in families, nursing homes, schools, or on cruise ships, or after ingestion of undercooked shellfish</td>
<td>Fever: Variable; Abdominal Pain: Common; Bloody Stool: NC; Vomiting, Nausea, or Both: NC; Fecal Evidence of Inflammation: NC; Hemepositive Stool: NC</td>
</tr>
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</table>

* If the illness can be linked to the ingestion of a particular food or meal, as in an outbreak, the incubation period can be deduced and the differential diagnosis narrowed accordingly. If the incubation period is less than 6 hours, *S. aureus* and *B. cereus* are likely causes; if the incubation period is 6 to 24 hours, *C. perfringens* and *B. cereus* are likely causes; and if the incubation period is 16 to 72 hours, possible causes include noroviruses, enterotoxigenic *E. coli*, vibrio, salmonella, shigella, campylobacter, yersinia, Shiga toxin–producing *E. coli*, giardia, cyclospora, and cryptosporidium. NC denotes not characteristic.
† Bloody diarrhea in an afebrile patient is suggestive of Shiga toxin–producing *E. coli* infection, although this is not always the case.
‡ Leukocytosis is found in approximately 50 percent of patients.
§ Classically, patients with intestinal amebiasis present with heme-positive or bloody stools.
¶ Although cryptosporidiosis is typically considered to be noninflammatory in the United States, fecal evidence of inflammation is common in children with cryptosporidiosis in developing countries.
¿ Fatigue, which may be profound, is reported in more than 90 percent of patients with cyclosporiasis.
**Vomiting occurs more frequently than diarrhea in children; diarrhea occurs more frequently in adults.
infection) and patients with severe, prolonged, or relapsing illness may benefit from antimicrobial therapy. Dramatic increases in the rate of fluoroquinolone resistance (contemporaneous with the widespread use of fluoroquinolones in poultry feeds) now limit the usefulness of these agents in campylobacter infections.

In uncomplicated salmonella gastroenteritis, antibiotics are not recommended, since they typically do not alleviate the diarrhea and have been associated with prolonged carriage and even relapse. However, because bacteremia occurs in 2 to 4 percent of patients, persons who are at increased risk for dissemination or metastatic seeding should receive antibiotics. These patients include those 12 months of age or younger and those older than 50 years; those with lymphoproliferative disorders, cancer, hemoglobinopathies, or AIDS; transplant recipients; patients with vascular grafts, artificial joints, degenerative joint diseases, or valvular heart disease; and patients who are taking corticosteroids.

Clues to the diagnosis of Shiga toxin–producing E. coli infection include bloody diarrhea (especially if the patient is afebrile) and a history of eating rare hamburger meat or seed sprouts. Given the concern that antimicrobial agents such as trimethoprim–sulfamethoxazole and fluoroquinolones induce

### Table 2. Antimicrobial Recommendations for Infections with Specific Pathogens

<table>
<thead>
<tr>
<th>Organism</th>
<th>Recommendations for Adults</th>
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<tbody>
<tr>
<td>Shigella</td>
<td>Fluoroquinolones (e.g., treatment for 1–3 days with ciprofloxacin, 500 mg orally twice daily; norfloxacin, 400 mg orally twice daily; or levofloxacin, 500 mg orally once daily); if susceptible, trimethoprim–sulfamethoxazole, 160 mg and 800 mg, respectively, twice a day for 3 days may be used.</td>
</tr>
<tr>
<td>Nontyphi species of salmonella</td>
<td>Not recommended in healthy host with mild or moderate symptoms, but if illness is severe or patient &lt;12 mo of age or &gt;50 yr of age or with prostheses, valvular heart disease, severe atherosclerosis, cancer, or uremia, treatment with trimethoprim–sulfamethoxazole (if susceptible) or fluoroquinolone, as above, for 5–7 days, or ceftriaxone, 100 mg/kg of body weight/day in 1 or 2 divided doses.</td>
</tr>
<tr>
<td><em>E. coli</em> (enterotoxigenic, enteropathogenic, or enteroinvasive)</td>
<td>Fluoroquinolones (e.g., treatment for 1–3 days with ciprofloxacin, 500 mg orally twice daily; norfloxacin, 400 mg orally twice daily; or levofloxacin, 500 mg orally once daily); if susceptible, treatment for 1–3 days with trimethoprim–sulfamethoxazole, 160 mg and 800 mg, respectively, twice daily may be used.</td>
</tr>
<tr>
<td><em>E. coli</em> Shiga toxin–producing (O157:H7)</td>
<td>Antimotility agents and antibiotics should be avoided, particularly trimethoprim–sulfamethoxazole and fluoroquinolones.</td>
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<tr>
<td>Yersinia</td>
<td>Antibiotics not usually required; for severe infections, doxycycline and aminoglycoside (in combination), trimethoprim–sulfamethoxazole, or fluoroquinolone therapy.</td>
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<tr>
<td><em>Vibrio cholerae</em> O1 or O139</td>
<td>Treatment with a single dose of doxycycline, 300 mg; treatment for 3 days with tetracycline, 500 mg 4 times daily; or single-dose treatment with fluoroquinolone.†</td>
</tr>
<tr>
<td>Toxigenic <em>C. difficile</em></td>
<td>Treatment with any offending antimicrobial agents should be discontinued, if possible; treatment for 10 days with metronidazole, 250 mg 4 times daily to 500 mg 3 times daily.</td>
</tr>
<tr>
<td>Giardia</td>
<td>Treatment for 7–10 days with metronidazole, 250 to 750 mg 3 times daily.</td>
</tr>
<tr>
<td>Cryptosporidium</td>
<td>If illness is severe or patient is immunocompromised, either paromomycin plus azithromycin or nitazoxanide should be considered; in patients with AIDS, highly active antiretroviral therapy sufficient to achieve immunologic reconstitution is most effective; in immunocompetent children with cryptosporidiosis, nitazoxanide should be considered.</td>
</tr>
<tr>
<td>Isospora</td>
<td>Treatment for 7–10 days with trimethoprim–sulfamethoxazole, 160 mg and 800 mg, respectively, twice daily.</td>
</tr>
<tr>
<td>Cyclospora</td>
<td>Treatment for 7–10 days with trimethoprim–sulfamethoxazole, 160 mg and 800 mg, respectively, twice daily.</td>
</tr>
<tr>
<td>Microsporidia</td>
<td>Treatment for 3 wk with albendazole, 400 mg twice daily; highly active antiretroviral therapy sufficient to achieve immunologic reconstitution is effective in patients with AIDS.</td>
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<tr>
<td><em>E. histolytica</em></td>
<td>Treatment for 5–10 days with metronidazole, 750 mg 3 times daily, plus either treatment for 20 days with iodoquinol, 650 mg 3 times daily, or treatment for 7 days with paromomycin, 500 mg 3 times daily.</td>
</tr>
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</table>

* There is a growing body of evidence suggesting that the use of fluoroquinolones is safe in children, but they should be used with caution in children, given reports of damage to articular cartilage of young beagles in preclinical testing.† Doxycycline and tetracycline may cause permanent discoloration of teeth during tooth development and are not recommended for use in children younger than 8 years of age.
Shiga-toxin production\textsuperscript{31} and may increase the risk of the hemolytic–uremic syndrome,\textsuperscript{30} patients with suspected or documented infection with Shiga toxin–producing E. coli should receive supportive care. Risk factors for the development of the hemolytic–uremic syndrome with this infection remain controversial and are incompletely understood.\textsuperscript{41}

**EMPIRICAL ANTIBIOTIC TREATMENT FOR COMMUNITY-ACQUIRED ILLNESS**

Empirical antibiotic treatment in adults who present with severe, community-acquired diarrhea (Table 3) reduces the average duration of illness by one to two days.\textsuperscript{28,42,43} However, as noted above, the potential benefits must be weighed against the potential harm, such as prolonged fecal shedding of certain pathogens, the increased risk of relapse, and the increased risk of complications. In patients who have evidence of inflammatory diarrhea (fever, tenesmus, dysentery, or fecal leukocytes or lactoferrin) and in whom diarrhea is not thought to be attributable to Shiga toxin–producing E. coli or fluoroquinolone-resistant campylobacter, empirical treatment with an agent such as a fluoroquinolone antibiotic (or for children, trimethoprim–sulfamethoxazole), pending fecal testing, is reasonable.\textsuperscript{6,44}

Given the increasing incidence of fluoroquinolone-resistant campylobacter,\textsuperscript{38,45} empirical therapy in immunocompromised or severely ill patients or travelers with inflammatory diarrhea (especially those who have traveled to southern Asia, where the rate of quinolone resistance exceeds 80 percent) might include erythromycin or azithromycin.

**TRAVELER’S DIARRHEA**

Although most cases of traveler’s diarrhea are self-limited, several clinical trials have demonstrated that fluoroquinolones or other agents — many of which are no longer used because of increased resistance — reduce the duration of diarrhea from three or four days to one or two days with as little as a single dose.\textsuperscript{46} Although such data support a recommendation that travelers carry appropriate antibiotics for the self-treatment of diarrhea, most experts do not recommend antimicrobial prophylaxis for travelers (even those who are immunocompromised, whose highest risk may be for infection with pathogens that are not covered by the standard antimicrobial agents used). Use of bismuth subsalicylate (one to two 262-mg tablets taken four times daily) decreases the incidence of traveler’s diarrhea by 35 to 65 percent and appears to be safe when used for up to three weeks but may result in the blackening of the tongue and the stools.\textsuperscript{47,48}

| Table 3. Considerations for Empirical Antimicrobial Therapy for Infectious Diarrhea.\textsuperscript{6} |
|---------------------------------|---------------------------------|---------------------------------|
| **Type of Illness**            | **Recommendations for Adults**  | **Comments and Alternatives**   |
| Moderate-to-severe traveler’s diarrhea | Treatment for 1–5 days with ciprofloxacin, 500 mg orally twice daily; norfloxacin, 400 mg orally twice daily; or levofloxacin, 500 mg orally once daily | Early treatment with a fluoroquinolone can reduce the duration of symptoms from 3–4 days to <1–2 days; in children, trimethoprim–sulfamethoxazole or a short course (1–3 days) of a fluoroquinolone should be considered. |
| Mild, community-acquired diarrhea, especially if invasive disease is suspected | Treatment for 1–5 days with ciprofloxacin, 500 mg orally twice daily; norfloxacin, 400 mg orally twice daily; or levofloxacin, 500 mg orally once daily | In children, trimethoprim–sulfamethoxazole or a fluoroquinolone should be considered; in immunocompromised patients, the addition of erythromycin or azithromycin should be considered for fluoroquinolone-resistant campylobacter infection; antimotility agents, quinolones, and trimethoprim–sulfamethoxazole should be avoided if infection with Shiga toxin–producing E. coli is suspected (e.g., if there is bloody diarrhea in an afebrile patient). |
| Severe nosocomial diarrhea, pending results of assay for C. difficile toxin | Treatment with any offending antibiotics should be discontinued if possible; treatment with metronidazole, 250 mg 4 times daily to 500 mg 3 times daily (continued for 10 days if assay for C. difficile toxin is positive) | Metronidazole therapy should be discontinued if assay for C. difficile toxin is negative. |
| Persistent diarrhea with suspected giardia infection | Treatment for 7–10 days with metronidazole, 250 to 750 mg 3 times daily | |

* There is a growing body of evidence suggesting that the use of fluoroquinolones is safe in children, but they should be used with caution in children, given reports of damage to articular cartilage of young beagles in preclinical testing.
**NOSOCOMIAL DIARRHEA**

The treatment of *C. difficile*–associated diarrhea has been discussed in detail previously. Many patients have improvement if therapy with broad-spectrum antibiotics can be discontinued. Oral metronidazole (which is less expensive) and oral vancomycin are effective in the treatment of *C. difficile*–associated diarrhea, with response rates of more than 90 percent and a mean time to improvement of three days, but relapses are common.

**PERSISTENT DIARRHEA**

The protozoal pathogens that are most frequently associated with diarrhea that persists for more than 7 to 10 days, both in the United States and worldwide, are *giardia* and *cryptosporidium*. Giardia infections typically respond to metronidazole (the use of larger doses, if they are tolerated, may result in fewer treatment failures) or tinidazole. Cryptosporidium infection, in contrast, is notoriously difficult to treat, particularly in patients with AIDS and low CD4+ lymphocyte counts, in whom the most effective strategy is to facilitate immunologic reconstitution with optimal antiretroviral therapy. Nizatidine, which was recently approved by the Food and Drug Administration (FDA) for use in children, may reduce oocyst shedding and diminish diarrhea. Since these hardy oocysts are difficult to filter or to kill with chlorine, infections caused by them are best prevented by heating water or food to near-boiling temperatures. Finally, cyclospora infection, which can be detected by means of acid-fast staining of fecal specimens, is readily treated with trimethoprim–sulfamethoxazole.

**AREAS OF UNCERTAINTY**

The inability to determine the specific causative agent when a patient is first seen greatly complicates management. Technology that will permit rapid, pathogen-specific diagnostic testing could mitigate the inappropriate use of antibiotics, help to track sources and outbreaks, and identify resistant pathogens or fecal contamination of food or water. Whether certain antibiotics increase risk whereas others prevent the hemolytic–uremic syndrome in patients with Shiga toxin–producing *E. coli* infection with quinolone-resistant *campylobacter*). One recent history of eating raw oysters to guide laboratory evaluation and management. For example, a recent history of eating raw oysters should prompt consideration and special culture for *vibrio*, and a recent history of antibiotic use should prompt testing for *Shiga* toxin–producing *E. coli*; the presence of bloody diarrhea would also warrant testing for shigella, salmonella, and campylobacter. A history of immunocompromise, sickle cell disease, severe atherosclerosis, or intravascular prosthetic material should prompt the testing of blood and stool cultures for salmonella. Recent travel to a developing country would suggest infection with pathogens that typically cause traveler’s diarrhea and would warrant empirical therapy with a single dose of a quinolone antibiotic (unless the travel was to southern Asia, which should arouse concern about infection with quinolone-resistant campylobacter). One might also consider empirical treatment for inflam-

**GUIDELINES**

Several groups have published guidelines for the management of acute diarrhea in adults. The guidelines that the Infectious Diseases Society of America published in 2001, with which we still concur (available at http://www.idsociety.org), detail diagnostic strategies, treatment recommendations, and public health implications for adults, listing the quality and strength of evidence supporting each recommendation. Several pediatric guidelines emphasize fluid management and the fact that antidiarrheal compounds are not useful.

**CONCLUSIONS AND RECOMMENDATIONS**

In healthy patients with an acute onset of diarrhea, such as the patient described in the vignette, epidemiologic and clinical evaluation should be used to guide laboratory evaluation and management. Whether certain antibiotics increase risk whereas others prevent the hemolytic–uremic syndrome in patients with Shiga toxin–producing *E. coli* infection with quinolone-resistant *campylobacter*). One recent history of eating raw oysters should prompt consideration and special culture for *vibrio*, and a recent history of antibiotic use should prompt testing for *Shiga* toxin–producing *E. coli*; the presence of bloody diarrhea would also warrant testing for shigella, salmonella, and campylobacter. A history of immunocompromise, sickle cell disease, severe atherosclerosis, or intravascular prosthetic material should prompt the testing of blood and stool cultures for salmonella. Recent travel to a developing country would suggest infection with pathogens that typically cause traveler’s diarrhea and would warrant empirical therapy with a single dose of a quinolone antibiotic (unless the travel was to southern Asia, which should arouse concern about infection with quinolone-resistant campylobacter). One might also consider empirical treatment for inflam-
matory diarrhea whose cause was suspected, on the basis of the exposure history, to shigella or campylobacter infection (Tables 2 and 3). If none of the above pathogens were present, but symptoms worsened or persisted for more than five days (especially if there was fever), we would order a fecal examination for inflammation to direct further testing for invasive pathogens (including Salmonella, shigella, campylobacter, C. difficile, or E. histolytica); if the examinations were negative for these pathogens, we would evaluate the patient for inflammatory bowel disease. Patients with illness lasting more than seven days should be tested for parasites (including giardia, cryptosporidium, cyclospora, and isospora). Finally, patients with recurrent or refractory giardiasis should be tested for possible immune compromise, such as immunoglobulin A deficiency, and patients with refractory cryptosporidiosis, cyclosporiasis, or isosporiasis should be tested for HIV infection.

Drs. Thielman and Guerrant are named as coinventors on a patent held by the University of Virginia for the use of glutamine derivatives in oral rehydration therapy. Dr. Guerrant is named as a coinventor on a patent held by the University of Virginia for a fecal lactoferrin assay. Neither coinventor receives any patent royalties.

References


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